Biodiversity of major bacterial groups in association with agarwood (Aquilaria crassna) in Khanh Hoa province, Vietnam

Da dạng sinh học các nhóm vi khuẩn chính trên Trâm hương Khánh Hòa, Việt Nam

Research article

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Agarwood mainly formed by Aquilaria species is an economically and pharmaceutically important natural product used for the production of incense, perfumes and traditional medicines in Asia. Endophytic bacteria are potentially important in producing pharmaceutical compounds found in the plants. The aim of this research is to isolate, classify and identify major endophytic bacteria groups associated with agarwood of Aquilaria crassna species in Khanh Hoa province, Vietnam. Agarwood samples were collected and surface-sterilized, and total endophytic bacteria were isolated on Tryptic Soy Agar by the spread plate method. Major bacterial groups were classified according to the Bergey’s system. The 16S rRNA gene fragments were amplified using PCR method, and bacterial isolates were identified using this gene sequence similarity based method. The results showed that from 0.121 g of agarwood, total 26 bacterial isolates were purified and divided into 7 separated groups, in which the group II of Gram-positive spore-forming bacteria was the most dominant. Especially, two dominant strains, T14 of group II, and T15 of group VII, were identified as Bacillus pumilus and Alcaligenes faecalis, respectively. To our knowledge, it is the first time that biodiversity of bacterial endophytes associated with agarwood from Aquilaria crassna in Vietnam has been reported, which requires further study to understand the relationship of endophytic bacteria to agarwood-producing Aquilaria crassna species as well as explore their potential applications towards the development of valuable bioactive compounds.

Keywords: agarwood, Aquilaria crassna, Bacillus pumilus, bioactive compounds, endophytic bacteria

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http://dx.doi.org/10.13141/jve.vol6.no2.pp132-137

ISSN: 2193-6471
1. Introduction

Agarwood (gaharu, jinko, aloeswood, oud, or tram huong in Vietnamese), which is the fragrant resinous wood, is an important commodity from *Aquilaria* and *Gyrinops* species in the family Thymelaeaceae and has been used as an incense, perfume, and multi-functional pharmaceutical in traditional medicine throughout Asia (Eurlings et al., 2010). Its pharmacological functions include sedative, analgesic, anti-inflammatory, anti-microbial, immunomodulatory, and wound healing properties. Also, agarwood is used as a digestive and laxative in medicine (Bhore et al., 2013; Huang et al., 2013). Due to increasing demand, agarwood has become very rare in the wild. Recently, all *Aquilaria* species are listed in Appendix II of the Convention on International Trade in Endangered Species of Wild Fauna and Flora (CITES) to improve control of commercial agarwood trade. *Aquilaria crassna* Pierre ex Lecomte, 1915, is the species that occurs in Vietnam, Cambodia, Laos, and central Thailand (Zhang et al., 2008).

Endophytes are microorganisms that live inside plants (inter or intra cellular in nature) without causing any plant disease (Bhore et al., 2013). Recent studies have indicated that endophytic bacteria are common among the resident microflora of the healthy inner tissues of various species of plants (Tian et al., 2007). Because the function of endophytic bacteria in the plants are poorly understood, isolation and identification of these bacterial strains can contribute into understanding the relationship between microbes and plants, especially agarwood-producing *Aquilaria* species, as well as the discovery of novel bacterial species. For example, a novel species *Micrococcus endophyticus* was uncovered from surface-sterilized *Aquilaria sinensis* roots (Chen et al., 2009). Recent results of 16S rRNA gene sequences analysis have revealed 18 different types of endophytic bacteria are associated with seven *Aquilaria* species in Malaysia (Bhore et al., 2013). In the present study, we report the results of isolation, primary classification and identification of major bacterial groups associated with agarwood produced by the economically and pharmaceutically important *Aquilaria crassna* species in Vietnam.

2. Materials and methods

2.1. Agarwood sampling and bacterial isolation

A total of 0.121 g of dark brown and soft agarwood (*Aquilaria crassna*) supported by Agarwood Society of Khanh Hoa province, Vietnam was used for bacterial isolation. The surface-sterilization of the collected agarwood samples was carried out as described elsewhere (Bhore et al., 2013). Total bacteria were isolated by the spread plate method. Agarwood samples were homogenized using tissue grinders and vortexed in sterile saline solution (8.5 g/1 NaCl). Tenfold serial dilution of samples was prepared and plated on Tryptic Soy Agar (TSA) (Difco, Detroit, MI, USA). All plates were incubated for 24–48 h at 37°C.

2.2. Morphological analysis and preliminary classification of bacteria isolates

Pure cultures of bacteria isolates were obtained by means of repeated streaking on plates containing TSA. The purified strains were picked and then maintained on TSA slants at 4°C and as 20% (w/v) glycerol suspensions at −70°C. Gram staining was carried out using the standard Gram reaction and was confirmed with the KOH test (Moaledj, 1986). Morphological analysis was carried out by using light microscopy (BH-2, Olympus) with cells from exponentially growing cultures. Colony morphology was observed on TSA after incubation at 37°C for 2-3 days.

Preliminary classification of bacteria isolates into major groups was performed according to *Berger's Manual of Determinative Bacteriology* (Holt, 1994).

2.3. Genomic DNA extraction and PCR amplification

Total DNA of bacterial isolates was extracted by the alkaline lysis method using the kit Wizard®SV Genomic DNA Purification System (Promega). Purified DNA samples were used as templates in the PCR reactions. The specific primers used for the amplification of the 16S rRNA gene (27F and 1492R) (Luan et al., 2007). The PCRs were performed in 50 µl reactions containing 2 µl (10 ng) of template DNA, 0.5 µM each primer, 1.5 mM MgCl2, 50 µM each dNTP, and 1 U Taq DNA polymerase along with 1X Taq buffer. Amplification was performed in a DNA thermal cycler (Bio-Rad Laboratories, Irvine, CA, USA) with an initial denaturing step for 10 min at 95°C and 40 cycles of 1 min per cycle at 95°C, 1 min at 55°C and 2 min at 72°C, followed by 5 min at 72°C. The amplified products were visualized in a 1% weight per volume (w/v) agarose gel (0.5 Trisborate-ethylene-diamine-tetraacetic acid buffer, pH 8.0) stained with ethidium bromide.

2.4. Gene sequencing and phylogenetic analyses

The PCR products of bacterial isolates were purified using the PCR Clean Up System Kit (Promega) and used as template for sequencing using the Big Dye terminator cycle sequencing kit (Applied Biosystems, Foster City, CA, USA) on an ABI 3130XL DNA Sequencer (Applied Biosystems) in Nam Khoa Biotek (Hochiminh city, Vietnam) and an ABI 3730XL DNA Sequencer (Applied Biosystems) in Marcrogen (Soul, Korea). Partial gene sequences of bacterial isolates and reference sequences available in GenBank were used for sequence analysis at the National Center for Biotechnology Information (NCBI) using BLAST (http://www.ncbi.nlm.nih.gov/BLAST). Gene sequences were aligned using ClustalW (Larkin et al., 2007), and regions with gaps were removed using BioEdit (Hall, 1999). Model selection was used to determine the best fit model with the lowest Bayesian Information Criterion.

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score for Neighbor Joining analysis, which was then used to construct a phylogenetic tree using the MEGA program ver. 5.2.1 (Tamura et al., 2011). The robustness of the tree topology was tested by bootstrap analysis with 1000 resamplings (Felsenstein, 1985).

3. Results

3.1. Isolation of bacteria in association with agarwood

The average total aerobic bacterial counts were recorded from the agarwood samples as $7.5 \times 10^4$ CFU/g. A total of 26 isolates were purified and named from T1 to T21 and T23 to T27.

3.2. Morphological diversity of bacterial isolates from agarwood

Colony and cell morphological characteristics of 26 isolates were illustrated in Figure 1 and Figure 2. Gram staining indicated that 16 isolates were Gram-positives and the remaining isolates were negatives (Figure 1). It is worth noting that more than a half of the number of Gram-positives including T7, T11, T12, T13, T14, T18, T21, T26 and T27 are endospore-forming bacteria, suggesting them as Bacillus or Clostridium.

The isolates T8, T9, T16, T20 and T23 indicated yellow colonies on TSA, whereas the T17 and T25 were green (Figure 2). All remaining isolates showed opalescent colonies. Interestingly, the isolates T17, T23, T24, T25 and T26 were found to modify the medium into green while yellow pigment was produced by the T27.

3.3. Classification of major bacterial groups isolated from agarwood

According to Bergey’s classification system, 26 isolates were divided into 7 different major groups of eubacteria (Table 1). The most dominant group is the group II with 9
isolates which indicate rod-shaped, spore-bearing, large and uniform cells of Bacillus and Clostridium.

Table 1. Major bacterial groups in association with agarwood (Aquilaria crassna) in Khanh Hoa province (Vietnam) according to the Bergey’s classification system

<table>
<thead>
<tr>
<th>Group</th>
<th>Strains</th>
<th>Gram staining</th>
<th>Cell characteristics</th>
<th>Endospore production</th>
<th>Representative genera in the Bergey’s system</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>T20</td>
<td>+</td>
<td>Round in clusters &amp; tetrads</td>
<td>-</td>
<td>Staphylococcus, Micrococcus, Peptococcus</td>
</tr>
<tr>
<td>II</td>
<td>T7, T11, T12, T13, T14, T18, T21, T26, T27</td>
<td>+</td>
<td>Rod-shaped, spore-bearing, large, uniform</td>
<td>+</td>
<td>Bacillus, Clostridium</td>
</tr>
<tr>
<td>III</td>
<td>T2, T8, T10, T17, T19, T23</td>
<td>+</td>
<td>Filamentous cell</td>
<td>-</td>
<td>Erysipelothrix, Lactobacillus, Eubacterium</td>
</tr>
<tr>
<td>IV</td>
<td>T4, T5, T16</td>
<td>-</td>
<td>Medium-sized coco-bacillary</td>
<td>-</td>
<td>Acinetobacter</td>
</tr>
<tr>
<td>V</td>
<td>T24, T25</td>
<td>-</td>
<td>Tiny coco-bacillary</td>
<td>-</td>
<td>Moraxella, Brucella, Bordetella, Bacteroides</td>
</tr>
<tr>
<td>VI</td>
<td>T6, T9</td>
<td>-</td>
<td>Pleomorphic cocccobacillary &amp; filamentous</td>
<td>-</td>
<td>Haemophilus, Bacteroides, Pasteurella, Francisella, Actinobacillus, Eikenella, Cardiobacterium, Flavobacterium</td>
</tr>
<tr>
<td>VII</td>
<td>T1, T3, T15</td>
<td>-</td>
<td>Uniformly Bacillary</td>
<td>-</td>
<td>Enterobacteriaceae, Pseudomonas, Aeromonas, Alcaligenes, Chromobacterium</td>
</tr>
</tbody>
</table>

3.4. Molecular identification of two dominant strains from agarwood

Two dominant strains (T14 and T15) were selected for further molecular identification into genus and species. The partial 16S rRNA gene of the strains T14 and T15 were sequenced with total 1340 and 1275 bp, respectively. BLAST analysis of the 16S rRNA gene sequence of T14 revealed 99.6-100% homology to type strains and others of the most related species including Bacillus pumilus, Bacillus altitudinis, Bacillus stratosphericus, Bacillus aerophilus, Bacillus altitudinis and Bacillus safensis. Phylogenetic analysis was shown in Figure 3, which confirmed this evolution relationship. Further analysis of biochemical characteristics using API 50 CHE kit identified the strain T14 as Bacillus pumilus (Table 2). Also, BLAST and phylogenetic analysis of the 16S rRNA gene sequence for the strain T15 showed 99.7% identity to the type strain NBRC 13111T of Alcaligenes faecalis and 99.1% identity to the type strain LMG 22996T of Alcaligenes aquatilis, which suggested T15 possibly belonged to Alcaligenes faecalis species (Figure 4).

Figure 3. Phylogenetic tree based on the 16S rRNA gene of the strain T14

Neighbor joining phylogenetic tree based on comparative analysis of 16S rRNA gene sequences of the strain T14 and the closest related type strains. Falsibacillus pallidus was used as an outgroup. GenBank accession numbers are shown in parentheses. Only branches with the percentage of replicate trees at least 50% in the bootstrap test (1000 replicates) was shown. The scale bar indicates the number of substitutions per nucleotide position.
Table 2. Differential phenotypic characteristics among the strain T14 and the related Bacillus species

<table>
<thead>
<tr>
<th>Carbohydrate metabolism</th>
<th>Strain T14</th>
<th>Bacillus pu- milis</th>
<th>B. safensis</th>
<th>B. altitudinis</th>
<th>B. aerophilus</th>
<th>B. stratosphercicus</th>
</tr>
</thead>
<tbody>
<tr>
<td>References</td>
<td><em>This study</em></td>
<td>API Satomi <em>et al.</em>, 2006</td>
<td>Shivaji <em>et al.</em>, 2006</td>
<td>Shivaji <em>et al.</em>, 2006</td>
<td>Shivaji <em>et al.</em>, 2006</td>
<td></td>
</tr>
<tr>
<td>Glycerol</td>
<td>w +/-</td>
<td>+/+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>D-arabinose</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>L-sorbose</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>L-rhamnose</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Inositol</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>+</td>
</tr>
<tr>
<td>Inulin</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>D-raffinose</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>D-cellubiose</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>+</td>
</tr>
<tr>
<td>D-trehalose</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

The scale bar indicates the number of substitutions per nucleotide position. 

**Figure 4. Phylogenetic tree based on the 16S rRNA gene of the strain T15**

Neighbor joining phylogenetic tree based on comparative analysis of 16S rRNA gene sequences of the strain T15 and the closest related strains. *Parapusillimonas granuli* was used as an outgroup. GenBank accession numbers are shown in parentheses. Only branches with the percentage of replicate trees at least 50% in the bootstrap test (1000 replicates) was shown. The scale bar indicates the number of substitutions per nucleotide position. T indicates type strains.

**4. Discussion**

Endophytes are endosymbiotic microbes, often bacteria or fungi, which live within a plant for at least part of its life without causing apparent disease in the plant (Bhore *et al.*, 2013). Endophytes have been found in all the species of plants studied to date; however, the relationship between endophytes and plants are not often well understood. Many fungal endophytes were shown to support the growth, enhance stress tolerance or improve animal resistance in the host plant (Cheplick and Faeth, 2009). Similarly, bacterial endophytes can produce bioactive compounds found in their host and have potential in providing new drugs, plant hormones, and novel natural products (Bhore *et al.*, 2013).

In *Aquilaria* species, agarwood was produced in response to wounding and fungal attack, so fungal endophytes were the most considered. Molecular methods revealed succession patterns of fungi associated with wound-induced agarwood in wild *Aquilaria malaccensis*, which include *Cunninghamella bainieri*, *Fusarium solani* and *Lasiodiplodia theobromae* (Mohamed *et al.*, 2014). Meanwhile, *Aspergillus phoenics*, *Penicilium citrum* and *Penicilium* spp. were considered as important fungi in the formation of agarwood in *Aquilaria crassna* in Vietnam.

To our knowledge, it is the first time that biodiversity of bacterial endophytes associated with agarwood from *Aquilaria crassna* in Vietnam has been reported. A total of 26 isolates of bacterial endophytes in *Aquilaria crassna*, which was composed of both Gram positives and negatives, both endospore and non-endospore-bearing forms, with diverse morphological characteristics in cells and colonies, was isolated and classified into seven different groups according to Bergey’s system. Especially, the group II with nine isolates included Gram-positive endospore-producing bacteria (e.g. *Bacillus* and *Clastridium*), which were the most dominant group. One of these isolates, T14, was identified as *Bacillus pumilus*, a Gram positive, aerobic, spore-forming bacillus commonly found in soil. Another soil bacterium, T15, was identified as *Alcaligenes faecalis*.

Interestingly, a recent study indicated that 13 out of 18 different types of endophytic bacteria associated with seven *Aquilaria* species in Malaysia belonged to *Bacillus* species and *Bacillus pumilus* was also found as the most common type (Bhore *et al.*, 2013). Many *Bacillus* species are found to secrete large quantities of different enzymes, so have become important species in many fields of the industry. Besides, *Bacillus* species can produce other natural products including bacteriocins and antibiotics, and therefore, they are potential sources of the novel natural products (Nguyen *et al.*, 2014). Other bacterial endophytes were also found to express their antimicrobial activities including antibacterial, antifungal, and antiviral function (Castillo *et al.*, 2002; Ding *et al.*, 2010, 2011). Therefore, further study is necessary to develop the potential applications of *Bacillus* species and other isolated bacterial endophytes. Also, uncovering the relationship between *Aquilaria* species and associated bacteria perhaps contribute into the protection of threatened *Aquilaria*
crassna species (Zhang et al., 2008), and even the deeper understanding mechanism of agarwood formation, which is now still being considered as a consequence of wounding and fungal attack only (Huang et al., 2013).

5. Conclusions

From the agarwood of Agaritaria crassna species in Vietnam, we purified 26 isolates belonging to 7 different groups of culturable bacterial endophytes. Among them, Gram-positive endospore-producing bacteria (e.g. Bacillus) were the most dominant group. Two strains T14 and T15 were identified as Bacillus pumilus and Alcaligenes faecalis, respectively. Further research is needed in order to understand the benefits of these bacterial endophytes to Agaritaria crassna species and look for economically and pharmaceutically important bioactive compounds from these producers.

6. References


