



RESEARCH ARTICLE

Efficacy of CaCl_2 against some important postharvest fungi on orange, chilli and Cavendish banana fruits

Hiệu quả của CaCl_2 đối với một số loại nấm quan trọng gây hại sau thu hoạch trên trái cam, ớt và chuối già

LE, Thanh Toan*; VO, Trong Ky; NGUYEN, Thi My Linh; TRIEU, Phuong Linh; NGO, Van Toan; NGUYEN, Huy Hoang

College of Agriculture and Applied Biology, Can Tho University, 3/2 Str., Can Tho city, Viet Nam

Fruit rot caused by *Aspergillus niger* or *Colletotrichum musae* is an important post-harvest disease on orange, chilli and Cavendish banana fruits. The use of synthetic fungicides has been a traditional strategy for the management of the fruit rot disease, but these chemicals adversely affect human health and environment. Therefore, the objective of this study was to evaluate the effects of CaCl_2 on *in vitro* hyphal growth and *in vivo* lesion inhibition. First, aqueous solutions of CaCl_2 at three concentrations of 20, 40 and 60 mM were assessed for their inhibitory effect against hyphal growth *in vitro*. Next, mature fruits were immersed into a solution of 20 mM CaCl_2 for 20 - 30 s, then inoculated by a pathogen suspension at the density of 10^6 conidia mL^{-1} and observed for 12 days. The results showed that 20 mM CaCl_2 was the most effective concentration in antifungal assay to *Aspergillus* isolated from orange rot. The treatment of CaCl_2 continued to gain efficacy on limiting lesions' development on orange fruits until 12 days after inoculation (DAI). On chilli, CaCl_2 at concentrations of 20 and 40 mM inhibited well on the growth of *Aspergillus* hyphae isolated from chilli rot. However, calcium treatment was not effective on chilli fruits. On Cavendish banana, solutions of CaCl_2 at concentrations of 20, 40 and 60 mM highly limited fungal growth of *Colletotrichum* *in vitro* conditions. The application of CaCl_2 solution could inhibit anthracnose lesion length of Cavendish banana variety, but its efficacy did not prolong until 6 DAI. In general, the good results were obtained from the 20 mM CaCl_2 in almost all the studied assays. Management of rot diseases on fruits by employing 20 mM CaCl_2 could be suitable to replace the current hazardous agro-chemicals.

Thối trái do nấm *Aspergillus niger* hay nấm *Colletotrichum musae* là bệnh sau thu hoạch thường gặp trên cam, ớt và chuối già. Thuốc trừ nấm tổng hợp là biện pháp truyền thống quản lý bệnh thối trái nhưng lại ảnh hưởng bất lợi đến sức khỏe con người và môi trường. Vì vậy, mục tiêu của nghiên cứu là đánh giá ảnh hưởng của CaCl_2 đối với sự sinh trưởng *in vitro* của nấm và sự ức chế vết bệnh ở điều kiện *in vivo*. Đầu tiên, dung dịch CaCl_2 ở các nồng độ 20, 40 và 60 mM được sử dụng để đánh giá khả năng ức chế sự sinh trưởng *in vitro* của nấm bệnh. Tiếp theo, trái trưởng thành được nhúng vào dung dịch CaCl_2 20 mM trong 20 - 30 s, rồi lây nhiễm với huyền phù mầm bệnh ở mật số 10^6 bào tử mL^{-1} và quan sát đến 12 ngày. Kết quả cho thấy CaCl_2 20 mM có hiệu quả ức chế tốt đối với nấm *Aspergillus* phân lập từ bệnh thối trái cam. CaCl_2 tiếp tục thể hiện hiệu quả ức chế bệnh trên trái cam đến 12 ngày sau lây bệnh. Trên ớt, CaCl_2 20 và 40 mM cho hiệu quả ức chế sự phát triển nấm *Aspergillus* phân lập từ bệnh thối trái ớt. Tuy nhiên, xử lý CaCl_2 không mang lại hiệu quả mong đợi trên trái ớt. Trên chuối già, dung dịch CaCl_2 ở các nồng độ 20, 40 và 60 mM ức chế tốt sợi nấm *Colletotrichum* trong điều kiện *in vitro*. Dung dịch canxi có thể ức chế vết bệnh thán thư trên chuối già, nhưng hiệu quả không kéo dài đến 6 ngày sau lây bệnh. Nhìn chung, các kết quả tốt đều đạt được khi xử lý bằng CaCl_2 20 mM ở hầu hết các thí nghiệm. Việc kiểm soát bệnh thối trái bằng CaCl_2 20 mM có thể thay thế cho hóa chất nông nghiệp độc hại hiện nay.

Keywords: *Aspergillus niger*, *Colletotrichum musae*, hyphal development, lesion inhibition

1. Introduction

The world fruit and vegetable production has quickly increased several folds over the last decades, applies to many kinds of the major crops including orange, chilli, banana, cucurbits, tomato, and cabbage, according to FAO reports (FAO, 2008). The rapid production of fruit and vegetable has occurred for several reasons. Nutritional and medicinal researches elucidate the value and role of these

foods in protecting human health. Fruits and vegetables provide essential nutrients as: carbohydrates, vitamins, minerals and fibres. Moreover, the current active global market and internationally intense import/export businesses make it possible for fresh agricultural products which cultivate in one part of the world to be quickly shipped and available to consumers anywhere else in the world. Increased production and business agreements always place a high pressure on farmers that they cannot

* Corresponding author
Email: lttan134@gmail.com

allow diseases to affect their harvest products. Fruit rot caused by *Aspergillus niger* Tiegh. or *Colletotrichum musae* (Berk. & Curtis) Arx. is the most important post-harvest disease on fruits (Sarkar, 2016; Dashora and Sharma, 2018; Lema et al., 2018). Average post-harvest losses to these fungi have been estimated at approximately 20% at developed countries, and up to 50% at developing countries (Janisiewicz and Korsten, 2002; Florkowski et al., 2009).

In Viet Nam, despite several advances of the production and disease management on fruits and vegetables, farmers often face many challenges (Hue and Nghiem, 2014). Fruit rot caused by *A. niger* plays a major role on losses of orange and chilli production from fields to storage houses (Long, 2012; Xuyen, 2012). *A. niger* also causes rot diseases on other fruits and vegetables such as mango, lemon, grapefruit, onion and garlic (Hocking, 2006; Liaquat et al., 2016). This fungi species not only cracks and causes damage on fruit epidermis but also secretes aflatoxins, a group of harmfully carcinogenic mycotoxins. These toxic compounds are heat-stable and are non-degradable by a variety of food processing procedures. Especially, low-level exposure to aflatoxins may lead to a suppression of the immune system and increase susceptibility to diseases in humans (Pestka and Bondy, 1994). In banana, anthracnose caused by *C. musae*, usually produces black and brown spots on banana peels, leading to low price and severe economic losses (Lassois et al., 2010). Furthermore, *C. musae* was reported as the popular post-harvest pathogenic fungus in Cavendish bananas in Viet Nam (Hang, 2012).

Many strategies are available for managing pathogens in fresh agricultural products. On the first strategy, application of waxes to fruit surfaces was researched. Waxing could improve the fruit appearance, reduces spoilage due to chilling injury, decreases the respiration and transpiration rates and protects fruit epidermis against infection by fungal pathogens. However, waxes could not improve the quality of fruits. Another major disadvantage of using wax for coating the fruits is the development of off-flavor (Sharma and Singh, 2000). In another strategy, UV-C light was applied for managing diseases on fruits and vegetables. The optimum dose and the time required to achieve maximum protection after UV-C treatment against plant pathogens vary depending on the nature of products (D'hallewin et al., 1999). Besides, fruit diseases could be managed using beneficial pathogens. Thanh et al. (2016) indicated that beneficial *Streptomyces fradiae* and *Bacillus polyfermenticus* were capable of inhibiting fungal growth of *Neoscytalidium dimitiatum* causing brown spot disease on dragon fruits in Vietnam. Efficacy of heat treatments and other disease management strategies has also been assessed (Palou et al., 2001; Awang et al., 2011; Mahmud et al., 2008; Ayon-Reyna et al., 2017; Netravati and Jagadeesh, 2018). The efficacy of different heat procedures (aqueous immersion of oranges in: water at up to 75 °C for 150 s; 2-4% sodium carbonate aqueous solution at 45 °C for 60 - 150 s, or 1-4% sodium bicarbonate aqueous solution at room temperature for 150 s followed by storage at 20 °C for 7 days) was determined in order to control blue mold

disease of oranges caused by *Penicillium italicum*. The incidence of green mold caused by *P. digitatum* was reduced to approximately 1-12%, whereas the untreated fruits were entirely (100%) infected by green mold disease (Palou et al., 2001). Last but not least, calcium treatments have been reported to retain fruit firmness in various agricultural products, including apple (Conway et al., 1994), custard apple (Netravati and Jagadeesh, 2018), peach (Manganaris et al., 2007; Sohail et al., 2015; Gayed et al., 2017), persimmon (Bagheri et al., 2015), pomegranate (Mirdehghan and Ghotbi, 2014), blackberry (Turmanidze et al., 2016), cape gooseberry (Reyes-Medina et al., 2017), tomato (Arthur et al., 2015) and lemon (Valero et al., 1998). Soaking the dragon fruits for 40 min in solutions of CaCl₂ significantly increased the fruit Ca content in the fruit peels, leading to reduce the size of anthracnose lesions (Awang et al., 2011). In mango, solutions containing various concentrations of CaCl₂ could delay the fruit ripening (Mahmud et al., 2008). However, the values of flavors and taste of the mango fruits at ripening were 4.0, 3.0, 2.5 and 2.25 at 2.5, 5.0, 7.5 and 10% CaCl₂, respectively. Treatment of CaCl₂ or combination of hot water – CaCl₂ could reduce mycelial growth and germination of *C. gloeosporioides* *in vitro* as well as delay anthracnose symptom on papaya fruits (Madani et al., 2016; Ayon-Reyna et al., 2017). In addition, CaCl₂ at a concentration of 4% could reduce the severity of infection from different fungi including *Alternaria alternata*, *Alternaria solani*, *A. niger*, *Botrytis cinerea*, *Fusarium solani* and *Rhizopus stolonifer* on guava fruits (Hassanein et al., 2018). On calcium treated-fruits, the association between firmness retention and reduced rot incidence suggests that calcium might affect both these processes simultaneously through its cellular role in strengthening fruit cell walls (Fallahi et al., 1997; Conway et al., 1999). The major advantage of calcium treatment on fruits is the safety to human health and avoidance of environmental pollutions.

Current researches on preventing *Aspergillus* fruit rot on orange and chilli fruits, as well as anthracnose on Cavendish banana fruits have not been carried out with CaCl₂ yet. Therefore, the objective of this study was to assess the efficacy of CaCl₂ treatment on growth of hyphae and on the diameter of rot lesions on fruits.

2. Materials and methods

2.1 Materials

2.1.1. Fungal strains and culturing

Two samples of virulent strains of *A. niger* isolated from orange and chilli fruits, and one virulent strain of *C. musae* isolated from banana anthracnose lesions were cultivated at Department of Plant Protection, College of Agriculture and Applied Biology, Can Tho University. The fungi *A. niger* and *C. musae* were isolated from infected fruits and identified by molecular analysis (Hang, 2012; Long, 2012; Xuyen, 2012). The fungi were prepared on Potato Dextrose Agar

(PDA) at pH 6.5 at about 10-12 days before conducting experiments. Medium of PDA was prepared followed the procedure of Atlas (2004). The fungi were incubated at 28 ± 2 °C with relative humidity approximately 90%.

2.1.2. Chemicals

$\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$ (Catalogue No. 1023820500, purity $\geq 95\%$, Merck, Germany) was provided from Department of Plant Protection, Can Tho University.

Prior to this research, different concentrations of $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$ had been quickly screened to evaluate their effects on spore germination of *A. niger* and *C. musae*. Subsequently, $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$ at 20, 40 and 60 mM were chosen for this research because it gave the high efficacy in inhibiting spore germination.

2.2. Assessment of CaCl_2 -efficacy on hyphal development of post-harvest fungi *in vitro* conditions

The experiments were carried out in completely randomized design (CRD), with four treatments including CaCl_2 at three different concentrations (20, 40, and 60 mM), and a water control treatment with six replications, one petri dish for each replication. Separated experiment was done with *A. niger* isolated from orange and chilli fruits, with *C. musae* of banana anthracnose. Experimental steps conducted were followed the procedure of Dhingra and Sinclair (1995), Mahmud et al. (2008), and Hajano et al. (2012).

The aqueous solutions of CaCl_2 was prepared according to treatment above description. To assure complete solubility, each sample of aqueous CaCl_2 was magnetically stirred for 30 min. The CaCl_2 solutions were filtered through Whatman papers with pore size of approximately 0.2 μm . The filtered solutions were then poured into the medium of PDA at 55-60 °C for 2 min and gently shaken. Approximately 10 mL of each obtained mixture medium was immediately poured into Petri dishes. After media solidification, a hyphal round slice of fungi at approximately 5 mm-diameter was placed at in the center of each petri dish (Dhingra and Sinclair, 1995). Diameter of fungal colony was measured at 48, 72 and 96 hours for experiments with *A. niger*, and at 24, 48 and 72 hours for *C. musae*.

Each experiment was performed 3 times. Based on results of *in vitro* experiments, an effective concentration of 20mM of CaCl_2 was chosen to conduct following experiments on orange, chilli and Cavendish banana fruits.

2.3. Assessment of treating CaCl_2 at an effective concentration on fruits before inoculation with fungal suspension

The experiment was done in CRD with two treatments including CaCl_2 -treated and control treatments, with 12 replications, one fruit per one replication, four inoculated points per one orange fruit, one inoculated point per one chilli or banana fruit.

Experimental fruits including orange, chilli and Cavendish banana were chosen, and surface-disinfected with 95% ethanol (v/v) for one min, and washed two times with sterile distilled water to remove the alcohol residue. Next, the fruits were immersed in a solution of CaCl_2 for 20-30s, then, air-dried for 1-2 h at room temperature. On the untreated control, the fruits were handled identically, but sterile distilled water was used instead of CaCl_2 solution. After that, a bunch of sterile needles was used to create tiny holes with a depth of 2 mm on fruit epidermis, with four positions on orange fruits, one position on chilli or Cavendish banana fruits. One mL of fungal spore suspension at a density of 10^6 conidia mL^{-1} was dropped on these tiny holes.

Inoculated fruits were stored in an incubation chamber at 25 °C with relative humidity approximately 98% for 24 h. Finally, inoculated fruits were transferred into transparent nylon bags with wet cotton inside, at room temperature (Sivakumar et al., 2002; Cao et al., 2008; Talibi et al., 2011; Yu et al., 2012) to observe the disease symptoms.

Length of *Aspergillus* rot lesions was recorded at 8, 10 and 12 days after inoculation (DAI) (on orange fruits), 4, 6 and 8 DAI (on chilli fruits). Separated experiment on orange or chilli fruits was repeated three times.

Anthracnose lesions length on banana fruits were recorded at 5, 6 and 7 DAI (De Costa and Erabadupitiya, 2005). The experiment was done twice.

2.4. Statistical analysis

Data were subjected to an analysis of variance using SPSS 16.0 software package (IBM, USA). Individual comparisons between mean values were performed using Duncan's Multiple Range Test (DMRT) or t-test with a magnitude of p value at 0.05.

3. Results and discussion

3.1 Effect of CaCl_2 solutions against *Aspergillus niger* on orange fruits

3.1.1 Efficacy of CaCl_2 on *Aspergillus* hyphal development *in vitro*

Each tested concentration of CaCl_2 showed different fungal activity *in vitro* conditions (Table 1 and Figure 1). Results for the individual concentration varied during three observation time points. The optimal efficacy was observed using

the treatment with as solution of 20 mM CaCl₂. Colonial diameter of the treatment of 20 mM CaCl₂ was 78.2 mm, significantly lower than that of the control sample (85.3 mm) at 96 hours adding putting fungal slices.

Table 1. Efficacy of CaCl₂ on colonial diameter (mm) of *Aspergillus niger* in vitro

Treatment	Time after adding fungal slices (hours)		
	48 ^{1/}	72 ^{1/}	96 ^{1/}
20 mM CaCl ₂ ·2H ₂ O	40.7±5.8 ^c	61.4±7.2 ^c	78.2±6.9 ^b
40 mM CaCl ₂ ·2H ₂ O	48.0±6.1 ^b	70.8±7.1 ^b	88.0±6.4 ^a
60 mM CaCl ₂ ·2H ₂ O	56.1 ± 6.3 ^a	77.8±7.4 ^a	84.5±7.2 ^a
Water control	42.8 ± 5.6 ^c	70.3±6.5 ^b	85.3±7.3 ^a
Significance	*	*	*
Coefficient of variance (%)	7.26	5.38	5.41

^{1/} Mean ± SE (standard error) followed by the same letter do not differ significantly according to DMRT at p ≤ 0.05.

*: significant at p ≤ 0.05

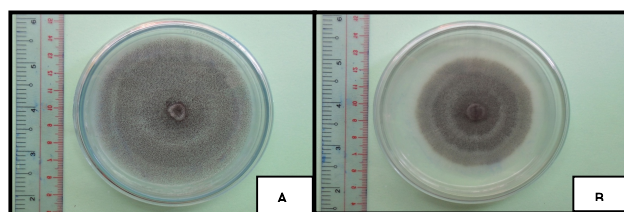


Figure 1. Efficacy of 20 mM CaCl₂ on the growth of *Aspergillus niger* at 96 hours after putting fungal slices.

The round slice of *Aspergillus* was put at the center of a Petri dish containing PDA medium and solution of CaCl₂ or distilled water. A: The control treatment with distilled water, B: the solution of CaCl₂ at a concentration of 20 mM.

3.1.2 Efficacy of treating CaCl₂ on oranges before an inoculation of *Aspergillus* suspension

Disease incidence on orange fruits was 100% (data not shown). The effect of CaCl₂ was assessed on the length of rot lesion on orange fruits at three observation time points. The lesion length of calcium treatment was significantly lower than that of the control at 8 DAI, and prolonged until 12 DAI (Table 2 and Figure 2).

The orange fruits were immersed on a solution of 20 mM CaCl₂ first, made tiny holes on orange epidermis at four positions by sterile needles, then inoculated with *Aspergillus* spore suspension at a density of 10⁶ conidia mL⁻¹. Control: the orange fruit was treated with sterile distilled water, CaCl₂ 20 mM: the orange fruit was treated with CaCl₂-solution at a concentration of 20 mM.

Table 2. Efficacy of CaCl₂ treated before *Aspergillus* inoculation on orange fruits

Treatment	Days after inoculation		
	8 ^{1/}	10 ^{1/}	12 ^{1/}
20 mM CaCl ₂ ·2H ₂ O	81.92±19.6 ^b	96.17±17.3 ^b	127.83±18.2 ^b
Non-treated control	105.5±19.8 ^a	131.17±15.3 ^a	161.00±15.3 ^a
Significance	*	*	*
Coefficient of variance (%)	27.99	27.17	21.57

^{1/} Mean ± SE (standard error) followed by the same letter do not differ significantly according to DMRT at p ≤ 0.05.

*: significant at p ≤ 0.05

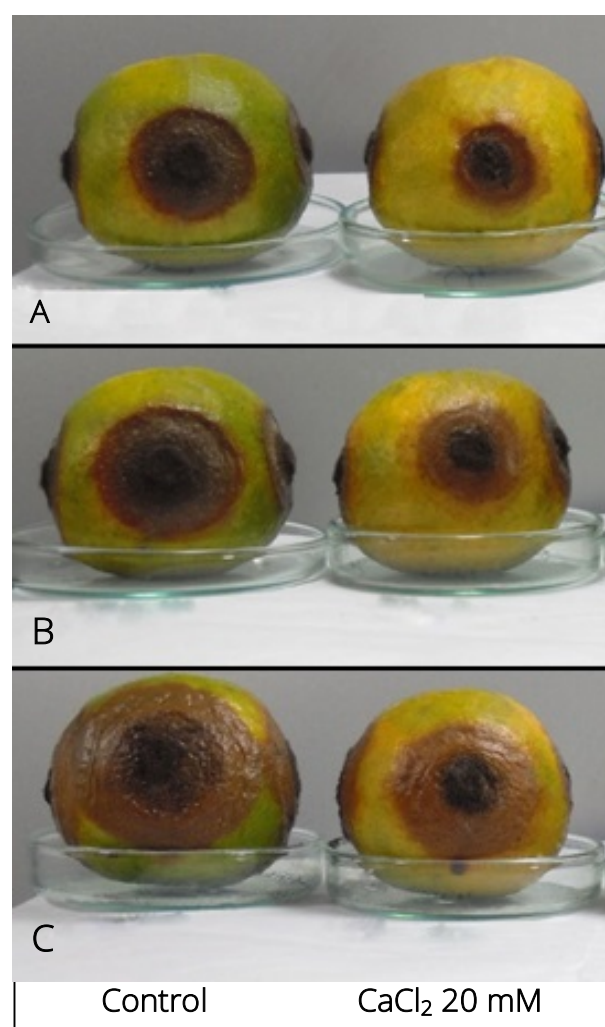


Figure 2. Efficacy of CaCl₂ at a concentration of 20 mM on rot lesions caused by *Aspergillus niger* at 8 DAI (A), 10 DAI (B), 12 DAI (C).

3.2 Effect of CaCl₂ solutions against *Aspergillus niger* on chilli fruits

3.2.1 Efficacy of CaCl₂ on *Aspergillus* hyphal development in vitro

The results showed that calcium treatment at a concentration of 20 mM CaCl₂ inhibited hyphal development of *A. niger* during all observation time points. The treatment of 20 mM CaCl₂ had ability from 48 to 96 hours after putting fungal slices, while CaCl₂ at a concentration of 60mM was effective on 72 and 96 hours after putting fungal slice. Therefore, the concentration at 20 mM of CaCl₂ showed a high and stable efficacy than other treatments, and was chosen to do on the following experiment (Table 3).

Table 3. Efficacy of CaCl₂ on colonial diameter (mm) of *Aspergillus niger* in vitro

Treatment	Time after adding fungal slices (hours)		
	48 ^{1/}	72 ^{1/}	96 ^{1/}
20 mM CaCl ₂ ·2H ₂ O	36.80±4.5 ^b	52.50±6.1 ^b	58.20±6.2 ^b
40 mM CaCl ₂ ·2H ₂ O	40.50±5.1 ^a	54.00±5.8 ^b	62.80±7.2 ^b
60 mM CaCl ₂ ·2H ₂ O	42.00±5.3 ^a	59.80±5.2 ^a	74.00±7.1 ^a
Water control	41.20±4.6 ^a	57.50±5.4 ^a	75.50±8.2 ^a
Significance	*	*	*
Coefficient of variance (%)	7.82%	5.85%	10.32%

^{1/} Mean ± SE (standard error) followed by the same letter do not differ significantly according to DMRT at p ≤ 0.05.

*: significant at p ≤ 0.05

3.2.2 Efficacy of treating CaCl₂ on chilli before an inoculation of *Aspergillus* suspension

Treatment of CaCl₂ at a concentration of 20 mM did not inhibit *Aspergillus* rot lesion on chilli fruits (Table 4).

Table 4. Efficacy of CaCl₂ treated before *Aspergillus* inoculation on chilli fruits

Treatment	Days after inoculation		
	4 ^{1/}	6 ^{1/}	8 ^{1/}
20 mM CaCl ₂ ·2H ₂ O	13.51±0.23	17.33±0.32	20.58±0.24
Non-treated control	10.34±0.19	16.76±0.33	19.02±0.27
Significance	ns	ns	ns
Coefficient of variance (%)	15.47%	21.78%	17.59%

^{1/} Mean ± SE (standard error) followed by the same letter do not differ significantly according to DMRT at P = 0.05.

*: significant at p ≤ 0.05; ns: non-significant at p ≤ 0.05

3.3 Effect of CaCl₂ solutions against *Colletotrichum musae* on Cavendish banana fruits

3.3.1 Efficacy of CaCl₂ on *Colletotrichum* hyphal development in vitro

Efficacy of CaCl₂ was determined by colonial diameters of *Colletotrichum musae* during three observation time points at 24, 48 and 72 hours after in vitro culture. At the time point of 24 h, all treatments of three concentrations of CaCl₂ had small colonies ranged from 8.66 mm to 10.00 mm, significantly lower than that of the control treatment of approximately 12.50 mm. At two following observation time points of 48 and 72 h, only CaCl₂ at a concentration of 20 mM was effective on inhibiting development of *Colletotrichum* colony, compared to those of control one (Table 5).

Table 5. Colonial diameter (mm) of *Colletotrichum musae* in vitro condition

Treatment	Time after adding fungal slices (hours)		
	24 ^{1/}	48 ^{1/}	72 ^{1/}
20 mM CaCl ₂ ·2H ₂ O	8.66±2.3 ^c	29.67±4.5 ^b	58.50±12.1 ^c
40 mM CaCl ₂ ·2H ₂ O	10.00±2.7 ^b	33.83±4.8 ^a	63.67±11.3 ^a
60 mM CaCl ₂ ·2H ₂ O	9.17±1.9 ^{bc}	34.33±5.2 ^a	63.17±9.3 ^{ab}
Water control	12.50±1.8 ^a	34.00±5.7 ^a	60.00±11.2 ^b
Significance	*	*	*
Coefficient of variance (%)	12.41	5.63	5.79

^{1/} Mean ± SE (standard error) followed by the same letter do not differ significantly according to DMRT at p ≤ 0.05.

*: significant at p ≤ 0.05

The effective concentration of CaCl₂ at 20 mM was chosen to carry out assays on Cavendish banana fruits.

3.3.2 Efficacy of treating CaCl₂ on Cavendish bananas before an inoculation of *Colletotrichum*

Lesion lengths of CaCl₂-treatment were short at approximately 11.92 mm, and statistically significant to those of control at 13.00 mm at 5 DAI. However, efficacy of CaCl₂ did not prolong to 6 and 7 DAI (Table 6 and Figure 3).

Table 6. Efficacy of CaCl₂ treated before *Colletotrichum* inoculation on Cavendish banana fruits

Treatment	Days after inoculation		
	5 ^{1/}	6 ^{1/}	7 ^{1/}
20 mM CaCl ₂ ·2H ₂ O	11.92±3.4 ^b	17.42±3.5	23.67±4.2
Non-treated control	13.00±3.6 ^a	18.25±3.2	25.08±3.8
Significance	*	ns	ns
Coefficient of variance (%)	11.12	12.22	11.53

¹Mean \pm SE (standard error) followed by the same letter do not differ significantly according to DMRT at $p \leq 0.05$; *: significant at $p \leq 0.05$; ns: non-significant at $p \leq 0.05$



Figure 3. Efficacy of CaCl₂ treatment before *Colletotrichum* inoculation on Cavendish bananas at 7 DAI.

The banana fruits of Cavendish were immersed on a solution of 20 mM CaCl₂ at approximately 20 s, air-dried for 2 h. Tiny holes were created by sterile needles with a depth of 2 mm on banana epidermis. One mL of *Colletotrichum* spore suspension at a density of 10⁶ conidia mL⁻¹ was dropped on these tiny holes. CaCl₂: the banana fruit was treated with CaCl₂ at a concentration of 20 mM, Control: the banana fruit was treated with distilled water.

4. Discussion

At *in vitro* conditions, the results showed that 20 mM CaCl₂ significantly inhibited the growth of the pathogenic fungi *A. niger* and *C. musae*, better than CaCl₂ solutions at 40 and 60 mM. Calcium solution at an optimal concentration was required for the inhibition of hyphae, whereas low or high concentrations could non-effect on hyphal growth. Fungal cells exposed to the optimal concentration increased their calcium level in the cytosol, leading to alter the osmotic balance and be toxic to the fungal cells (Al-Eryani-Raqeeb et al., 2009). The cell walls of fungal pathogens were rich in neutral sugars. When a low concentration of CaCl₂ was applied *in vitro* conditions, exogenous Ca²⁺ stimulated a production of glucosamine, a kind of amino sugars. The glucosamine is beneficial to fungal growth, resulted in failure on fungal inhibition. On the contrary, high calcium concentration reduced neutral sugars, but lead to an increase in uronic acid, Ca, P and Na in fungal cytosol. Uronic acid is a part of fungal cell walls. Besides, elements of Ca, P and Na play important roles in metabolism and energy transfer as well as an integral component of DNA, RNA, coenzymes and membrane phospholipids of fungal cells. Therefore,

fungal pathogens could grow in the PDA medium containing a high calcium concentration (Chardonnet et al., 1999; Stosic et al., 2014).

At *in vivo* conditions, the results showed that treatment of 20 mM CaCl₂ on orange, chilli and Cavendish banana reduced the lesions length. The reduction of rot symptom in CaCl₂-treated fruits could be due to calcium content of fruit peels and host-pathogen interaction. On fruits, calcium is mainly associated with the pectic materials. The Ca²⁺ ions could interact with the anionic pectic polysaccharides, coordinating with the oxygen functions of two adjacent pectin chains, and cross-linking the chains (Rose et al., 2003). The calcium binding could reduce the accessibility of cell wall degrading enzymes from *Aspergillus* fungus to the substrates of fruits. In the research of Marcelle (1995), the authors indicated that calcium treatment could conserve fruit epidermis, inhibit a post-harvest respiratory peak and delay the ripen process of apple fruits. Moreover, CaCl₂ could be treated at 2-3 weeks before harvesting, leading to prolong shelf life of fruit (Sen et al., 2001). On mango and strawberry fruits, soaking or dipping fruits into the solution of CaCl₂ lead to decrease fruit rot (Uthaibutra et al., 1998; Lara et al., 2004). On red-flesh dragon fruits, CaCl₂ applied at various concentrations at 1, 2, 3 and 4 gL⁻¹ reduced the anthracnose severity. The concentration of CaCl₂ at 4 gL⁻¹ (approximately 27 mM) gave the best inhibition on anthracnose lesions (Awang et al., 2011). Our results of 20 mM CaCl₂ on this study are in line to results of Awang et al. (2011). Similarly, CaCl₂ was reported to inhibit the *in vitro* development of *C. gloeosporioides* in papaya fruits (Ayon-Reyna et al., 2017). In a recent study, Hassanein et al. (2018) indicated that 4% (w/v) CaCl₂ could use with gamma irradiation to inhibit fungal growth of *A. alternate*, *A. solani*, *A. niger*, *B. cinerea*, *F. solani* and *R. stolonifer* on guava fruits.

Moreover, host-pathogen interaction on CaCl₂-treated fruits could be an important mechanism. The fungal pathogen must first attack the fruit skin, where fungus secretes cell wall degrading enzymes, favouring the fungal infection (Chardonnet et al., 1999). However, the strengthening of the fruit cell walls by Ca²⁺ binding the polygalacturonase, reduce the susceptibility to fungal infection (Rose et al., 2003), leading to slow the development of rot lesions on CaCl₂-treated fruits. In general, CaCl₂ application in adequate amounts supplied helps to maintain orange, chilli and banana fruits' firmness, decrease on an incidence of fungal attack, leading to limit the diameter of rot lesions.

5. Conclusion

The concentration at 20 mM of CaCl₂ well inhibited the hyphal development of both *A. niger* and *C. musae* *in vitro*. However, this concentration of CaCl₂ was effective on inhibiting *Aspergillus* rot lesion only on orange fruits *in vivo* conditions. However, a treatment of 20 mM CaCl₂ on chilli or Cavendish banana fruits had a lower efficacy than on orange fruits. The immersion of orange fruits in a solution

of 20 mM might be a useful strategy to control fruit rot. Future studies are needed to formulate a commercial solution of CaCl₂.

6. References

- [1] Al-Eryani-Raqeeb, A., Mahmud, T.M.M., Omar, S.R.S., Zaki, A.R.M., Al-Eryani, A.R. 2009. Effects of calcium and chitosan treatments on controlling anthracnose and post-harvest quality of papaya (*Carica papaya* L.). International Journal of Agricultural Research 4: 53-68.
- [2] Arthur, E., Oduro, I., Kumah, P. 2015. Postharvest quality response of tomato (*Lycopersicon esculentum* Mill.) fruits to different concentrations of calcium chloride at different dip-times. American Journal of Food and Nutrition 5(1): 1-8.
- [3] Atlas, R.M. 2004. Handbook of microbiological media, 3rd edition. CRC Press Boca Raton London New York Washington, D.C., USA. 2051p.
- [4] Ayon-Reyna, L.E., Lopez-Valenzuela, J.A., Delgado-Vargas, F., Lopez-Lopez, M.E., Molina-Corral, F.J., Carrillo-Lopez, A., Vega-Garcia, M.O. 2017. Effect of the combination hot water – calcium chloride on the *in vitro* growth of *Colletotrichum gloeosporioides* and the post-harvest quality of infected papaya. The Plant Pathology Journal 33(6): 572-581.
- [5] Awang, Y., Ghani, M.A.A., Sijam, K., Mohamad, R.B. 2011. Effect of calcium chloride on anthracnose disease and postharvest quality of red-flesh dragon fruits (*Hylocereus polyrhizus*). African Journal of Microbiology Research 5(29): 5250-5259.
- [6] Bagheri, M., Esna-Ashari, M., Ershadi, A. 2015. Effect of postharvest calcium chloride treatment on the storage life and quality of persimmon fruits (*Diopyros kaki* Thunb.) cv. 'Karaij'. International Journal of Horticultural Science and Technology 2(1): 15-26.
- [7] Cao, S., Zheng, Y., Yang, Z., Tang, S., Jin, P., Wang, K., Wang, X. 2008. Effects of methyl jasmonate on the inhibition of *Colletotrichum acutatum* infection in loquat fruit and the possible mechanisms. Postharvest Biology and Technology 49: 301-307.
- [8] Chardonnet, C.O., Sams, C.E., Conway, W.S. 1999. Calcium effect on the mycelial cell walls of *Botrytis cinerea*. Phytochemistry 52: 967-973.
- [9] Conway, W.S., Janisiewicz, W.J., Klein, J.D., Sams, C.E. 1999. Strategy for combining heat treatment, calcium infiltration, and biocontrol to reduce postharvest decay of "Gala" apples. HortScience 34: 700-704.
- [10] Conway, W.S., Sams, C.E., Wang, C.Y., Abbott, J.A. 1994. Additive effects of postharvest calcium and heat treatment on reducing decay and maintaining quality in apples. Journal of the American Society for Horticultural Science 119: 49-53.
- [11] D'hallewin, G., Schirra, M., Manueddu, E., Piga, A., Ben-Yehoshua, S. 1999. Scoparone and scopoletin accumulation and ultraviolet-C induced resistance to postharvest decay in oranges as influenced by harvest date. Journal of the American Society for Horticultural Science 124: 702-707.
- [12] Dashora, A., Sharma, K. 2018. Isolation of fungal species from infected chilli fruit and their identification. World Journal of Pharmacy and Pharmaceutical Sciences 7(8): 1146-1159.
- [13] De Costa, D.M., Erabadupitiya, H.R.U.T. 2005. An integrated method to control postharvest diseases of banana using a member of the *Burkholderia cepacia* complex. Postharvest Biology and Technology 36: 31-39.
- [14] Dhinggra, O.D., Sinclair, J.B. 1995. Basic plant pathology methods, 2nd edition. CRC Press. 434p.
- [15] Fallahi, E., Conway, W.S., Hickey, K.D., Sams, C.E. 1997. The role of calcium and nitrogen in postharvest quality and disease resistance of apples. HortScience 32: 831-835.
- [16] FAO (Food and Agriculture Organization of the United Nations). 2008. FAOStat. Retrieved from <http://www.fao.org/faostat>.
- [17] Florkowski, W.J., Shewfelt, R., Brueckner, B., Prussia, S.E. 2009. Postharvest handling – A systems approach. Academic Press, 30 Corporate Drive, Suite 400, Burlington, MA 01803, USA. 615p.
- [18] Gayed, A.A.N.A., Shaarawi, S.A.M.A., Elkhishen, M.A., Elsherbini, N.R.M. 2017. Pre-harvest application of calcium chloride and chitosan on fruit quality and storability of 'Early Swelling' peach during cold storage. Ciencia e Agrotecnologia 41(2): 220-223.
- [19] Hajano, J., Lodhi, A.M., Pathan, M.A., Khanzada, M.A., Shah, G.S. 2012. *In-vitro* evaluation of fungicides, plant extracts and bio-control agents against rice blast pathogen *Magnaporthe oryzae* Couch. Pakistan Journal Botany 44(5): 1775-1778.
- [20] Hang, L.T.T. 2012. Diagnosis of post-harvest fungal pathogens on banana fruits (*Musa sapientum* L.). Thesis in Plant Protection. Can Tho University. 50p.
- [21] Hassanein, R.A., Salem, E.A., Zahran, A.A. 2018. Efficacy of coupling gamma irradiation with calcium chloride and lemongrass oil in maintaining guava fruit quality and inhibiting fungal growth during cold storage. Folia Horticulture 30(1): 67-78.
- [22] Hocking, A.D. 2006. *Aspergillus* and related teleomorphs. In: Food spoilage microorganisms, Blackburn, C. de W. (Ed). Woodhead Publishing Ltd., Cambridge CB1 6AH, UK. pp 451-487.
- [23] Hue, N.T., Nghiem, H.D. 2014. Research on the status of chemical pesticide uses in agricultural activities in Phan Me commune, Phu Luong district, Thai Nguyen province. Journal of Vietnamese Environment 6(2):

- 138-141.
- [24] Janisiewicz, W.J., Korsten, L. 2002. Biological control of postharvest diseases of fruits. *Annual Review of Phytopathology* 40: 411-441.
- [25] Lara, I., Garcisa, P., Vendrell, M. 2004. Modification in cell wall composition calcium-treated strawberry (*Fragaria x ananassa* Duch) fruit. *Postharvest Biology and Technology* 34: 331-39.
- [26] Lassois, L., Jijakli, M.H., Chillet, M., De Bellair, L. 2010. Crow rot of bananas. Preharvest factor involved in postharvest disease development and integrated control methods. *Plant Disease* 94(6): 648-658.
- [27] Lema, A.A., Mudansiru, A., Alexander, B.A., Sakinatu, M.J. 2018. Evaluation of fungal species isolated from three different varieties of pepper (*Capsicum chinense*, *C. frutescens* and *C. annum* L.) in Dutsin-ma, Katsina State. *Annals of Biological Sciences* 6(1): 13-17.
- [28] Liaquat, F., Arif, S., Ashraf, M., Chaudhary, H.J., Munis, M.F. 2016. *Aspergillus niger* causes fruit rot of lemon and grapefruit in Pakistan. *Plant Disease* 100(9): p1951.
- [29] Long, P.P. 2012. Diagnosis of post-harvest fungal pathogens in chilli (*Capsicum* spp.) fruits. Thesis in Plant Protection. Can Tho University. 59p.
- [30] Madani, B., Mirshekari, A., Sofo, A., Mohamed, M.T.M. 2016. Preharvest calcium applications improve post-harvest quality of papaya fruits (*Carica papaya* L. cv. Eksotika II). *Journal of Plant Nutrition* 39(10): 1483-1492.
- [31] Mahmud, T.M.M., Kadir, J., Rahman, R.A., Begum, M.M. 2008. Antimicrobial activities of chitosan and canxi chloride on in vitro growth of *Colletotrichum gloeosporioides* from papaya. *Pertanika Journal of Tropical Agricultural Science* 31(2): 223-232.
- [32] Manganaris, G.A., Vasilakakis, M., Diamantidis, G., Mignani, I. 2007. The effect of postharvest calcium application on tissue calcium concentration, quality attributes, incidence of flesh browning and cell wall physicochemical aspects of peach fruits. *Food Chemistry* 100: 1385-1392.
- [33] Marcelle, D.R. 1995. Mineral nutrition and fruit quality. *Acta-Horticulturae* 383: 219-225.
- [34] Mirdehghan, S.H., Ghotbi, F. 2014. Effects of salicylic acid, jasmonic acid, and calcium chloride on reducing chilling injury of pomegranate (*Punica granatum* L.) fruit. *Journal of Agricultural Science and Technology* 16: 163-173.
- [35] Netravati, Suresh, G.J., Jagadeesh, S.L. 2018. Calcium chloride and wax influences the post harvest behaviour of custard apple fruits. *Journal of Pharmacognosy and Phytochemistry* 7(2): 79-84.
- [36] Palou, L., Smilanick, J.L., Usall, J., Viñas, I. 2001. Control of postharvest blue and green molds of oranges by hot water sodium carbonate and sodium bicarbonate. *Plant Disease* 85: 371-376.
- [37] Pestka, J.J., Bondy, G.S. 1994. Mycotoxin-induced immune modulation. In: *Immunotoxicology and immunopharmacology*, Dean, J.H., Luster, M.I., Munson, A.E., Kimber, I. (Eds.). New York: Raven. pp. 163-182.
- [38] Reyes-Medina, A.J., Pinzon, E.H., Alvarez-Herrera, J.G. 2017. Effect of calcium chloride and refrigeration on the quality and organoleptic characteristics of cape gooseberry (*Physalis peruviana* L.). *Acta Agronomica* 66(1): 15-20.
- [39] Rose, J.K.C., Catalá, C., Gonzalez-Carranza, Z.H., Roberts, J.A. 2003. Cell wall disassembly. In: *The plant cell wall*, Rose, J.K.C. (ed.). Blackwell Publishing Ltd., Oxford, UK, pp. 264-324.
- [40] Sarkar, A.K. 2016. Anthracnose diseases of some common medicinally important fruit plants. *Journal of Medicinal Plants Studies* 4(3): 233-236.
- [41] Sen, F., Karakali, I., Yildiz, M., Kinay, P., Yildiz, F., Iqbal, N. 2001. Storage ability of Satsuma as affected by preharvest treatments. *Acta Horticultural (ISHS)* 553: 77-78.
- [42] Sharma, R.M., Singh, R.R. 2000. Harvesting, postharvest handling and physiology of fruits and vegetables. In: *Postharvest technology of fruits and vegetables*. Verma, L.R., Joshi, V.K. (eds), Indus Publishing Co., New Delhi, India, pp. 94-147.
- [43] Sivakumar, D., Hewarathgamagae, N.K., Wijeratnam, R.S.W., Wijesundera, R.L.C. 2002. Effect of ammonium carbonate and sodium bicarbonate on anthracnose of papaya. *Phytoparasitica* 30(5): 486-492.
- [44] Sohail, M., Ayub, M., Khalil, S.A., Zeb, A., Ullah, F., Afridi, S.R., Ullah, R. 2015. Effect of calcium chloride treatment on post harvest quality of peach fruit during cold storage. *International Food Research Journal* 22(6): 2225-2229.
- [45] Stosic, S., Stojanovic, S., Milosavjevic, A., Dolovac, E.P., Zivkovic, S. 2014. Effect of calcium salts on postharvest fungal pathogens *in vitro*. *Plant Protection* 65(1): 40-46.
- [46] Talibi, I., Askarne, L., Boubaker, H., Boudyach, E.H., Msanda, F., Saadi, B., Aoumar, A.A.B. 2011. Antifungal activity of some Moroccan plants against *Geotrichum candidum*, the causal agent of postharvest citrus sour rot. *Journal of Crop Protection* 35: 41-46.
- [47] Thanh, L.H., Tam, N.K.B., Nga, V.T., Thuy, H.T., Van, T.H., Son, H.T., Quynh, N.N., Nga, N.T.H. 2016. Study on the possibility of using microorganisms as biological control fungal pathogens *Neoscytalidium dimidiatum* causing disease of brown spots on the dragon fruit. *Journal of Vietnamese Environment* 8(1): 41-44.
- [48] Turmanidze, T., Gulua, L., Jgenti, M., Wicker, L. 2016. Effect of calcium chloride treatments on quality char-

- acteristics of blackberry fruit during storage. International Journal of Food and Allied Sciences 2(2): 36-61.
- [49] Uthaibutra, J., Saenghn, K., Sornsrivichai, J., Kumpoun, W., Sardud, V. 1998. Effects of fruit position and pre-harvest calcium dips on "Nam Doc Mai" mango fruit quality. In: Disease control and storage life extension in fruit. Coates, L.M., Hofman, P.J., Johnson, G.I. Canberra, ACIAR Proceedings 81: 27-30.
- [50] Valero, D., Martínez-Romero, D., Serrano, M., Riquelme, F. 1998. Influence of postharvest treatment with putrescine and calcium on endogenous polyamines, firmness, and abscisic acid in lemon (*Citrus limon* L. Burn cv. Verna). Journal of Agricultural and Food Chemistry 46: 2102-2109.
- [51] Xuyen, N.T.M. 2012. Diagnosis of post-harvest fungal pathogens on orange cv. Sanh (*Citrus nobilis* var. *typica* Hassk) and mandarin cv. Duong (*Citrus reticulata* Blanco). Thesis on Plant Protection. Can Tho University. 55p.
- [52] Yu, T., Yu, C., Chen, F., Sheng, K., Zhou, T., Zunun, M., Abudu, O., Yang, S. 2012. Integrated control of blue mold in pear fruit by combined application of chitosan, a biocontrol yeast and calcium chloride. Journal of Postharvest Biology and Technology 69: 49-53.